CHS-114, an Anti-CCR8 Cytolytic Monoclonal Antibody That Preferentially Depletes Intratumoral Tregs and Induces Antitumor Immune Responses

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Introduction

- FOXP3⁺ regulatory T cells (Tregs) play a crucial role in modulating immune responses across several tissues, including the tumor microenvironment (TME).^{1,2}
- Broad depletion of Tregs may elicit deleterious autoimmunity; however, selective depletion of Tregs within the TME may enhance antitumor immune responses.
- CCR8 is a G protein-coupled chemokine receptor (GPCR) that is predominantly upregulated on tumor-resident Tregs. Preferential depletion of Tregs in the TME may be achieved through targeting of CCR8.^{3,}
- CHS-114 is a specific, highly selective, effector-enhanced, human anti-CCR8 afucosylated mAb that exhibits no cross-reactivity to CCR8 expressed in other species.
- CHS-114 is currently in Phase 1 clinical studies for advanced cancer patients (NCT05635643).







(A) CCR8⁺ cell frequency in immune cell subsets as determined by flow cytometry from healthy PBMC (n = 14) and dissociated primary tumor tissues (n = 55). Error = SD. (B) Multiplex immunofluorescence (mIF) evaluating FOXP3 and CCR8 shows abundant CCR8⁺ Tregs in the TME of an HNSCC tumor sample. Representative images at 10x (H&E and low-power mIF) and 20x magnification (high-power mIF). **(C)** The density of CCR8⁺ Tregs in 12 types of solid tumors was determined by mIF staining.





(A) Antibody specificity screen evaluating binding to 5,528 extracellular protein targets by cell microarray demonstrates that CHS-114 has no off-target binding compared to other CCR8 antibody candidates (includes evaluation of 3 clinical-stage CCR8 mAbs derived from patent sequences). (B) To assess binding affinity of CHS-114 to human CCR8, engineered human CCR8-expressing 293T cells were incubated with CHS-114 and measured by flow cytometry (FC). (C) To assess CHS-114-mediated ADCC, labeled Raji-hCCR8 cells were co-cultured with NK cells and treated with CHS-114 or isotype control for 4 hours and then analyzed by FC. % NK killing represents the frequency of viable dye+CFSE+Raji-hCCR8 cells among total CFSE⁺Raji-hCCR8 cells. (D) To assess CHS-114-mediated ADCP, CFSE-labeled Raji-CCR8 cells were co-cultured with monocyte-derived macrophages and were treated with CHS-114 or isotype control for 3 hours and then analyzed by FC. % phagocytosis represents the frequency of CD14⁺CFSE⁺ macrophages among total CD14⁺ macrophages. Error = SD.



(A) Schematic representation of the experimental system; dissociated human tumor cells (DTC; 3 RCC, 1 CRC, n = 4) were co-cultured with IL-2 activated allogeneic human NK cells, treated with CHS-114 or isotype control overnight, and analyzed by flow cytometry. (B) % remaining Tregs, CD4⁺ Tconv cells, and CD8⁺ T cells were calculated by dividing the frequency of CHS-114 (10 µg/mL) treated cells by the frequency of isotype-treated cells. (C) NK and (D) monocyte activation in DTC (RCC) treated with CHS-114 or isotype control (representative results are shown). Error = SD.







CCR8⁺ Tregs in the PBMC were measured from whole blood using flow cytometry before CHS-114 treatment (predose) and at indicated timepoints throughout CHS-114 Cycle 1 for dose levels (DL)3-DL7. (A) CCR8+ Treg depletion (>85%) in the periphery was stable through Cycle 1 for all dose levels tested. Additionally, depletion was observed at DL3 (and higher doses), which was lower than predicted dose from *in vitro* modeling. (B) CHS-114 treatment led to a decrease in subset of Tregs, while preserving broader Treg and (C) non-Treg CD4⁺ T cell populations, confirming the specificity of CHS-114 for CCR8⁺ Tregs. Tregs were defined as CD127^{low} CD25^{high} cells within the CD3⁺ CD4⁺ T cell population. Data representative of the average per dose level (n = 2-3 participant samples per timepoint). Error = SEM.

Abbreviations: 2L = second line; ADCC = antibody-dependent cellular cytotoxicity; ADCP = antibody-dependent cellular phagocytosis; BOIN = Bayesian optimal interval; CCR8 = chemokine receptor 8; ccRCC = clear cell renal cell carcinoma; CFSE = carboxyfluorescein succinimidyl ester; CRC = colorectal cancer; CxDy = Cycle x Day y; DC = dendritic cell; DL = dose level; DLT = dose-limiting toxicities; DTC = dissociated tumor cells; FC = flow cytometry; gMFI = global mean fluorescence intensity; GPCR = G protein-coupled receptor; H&E = hematoxylin and eosin; HCC = hepatocellular carcinoma; HNSCC = head and neck squamous cell carcinoma; IFN = interferon; IL = interleukin; IO = immuno-oncology IQR = interguartile range; mAb = monoclonal antibody; mIF = multiplex immunofluorescence; MSD = Meso Scale Discovery; NK = natural killer; NSCLC = non-small cell lung cancer; PBMC = peripheral blood mononuclear cells; PD-1 = programmed cell death protein 1; QnW = every n weeks; RCC = renal cell carcinoma; Response Evaluation Criteria in Solid Tumours CC = squamous cell carcinoma; SD = standard deviation; SEM = standard error of the mean; Tconv = conventional T cell; TEAE = treatment-emergent adverse event; Teff = effector cell; TME = tumor microenvironment; TNF = tumor necrosis factor; Treg = regulatory T cell. References: 1) Plitas et al, Annu Rev Cancer Biol 2020;4:459. 2) Sakaguchi et al, Annu Rev Immunol 2020;38:541. 3) Plitas et al, Immunity 2016;45:1122. 4) Wang et al, Nat Immunol 2019;20:1220

Statistics: * = p < 0.05; ** = p < 0.01; *** = p < 0.001; **** = p < 0.0001; when not shown, comparisons are not statistically significant.



CHS-114 Antibody Concentrations in Cancer Patients

CHS-114 PK exposure increases with dose. Dose proportionality is observed at all dose levels. PK elimination is linear over all dosing levels with a half-life of approximately 10 days (range: 9-17 days).







CHS-114 treatment of cancer patients induces proinflammatory cytokines. Serum cytokines were measured by MSD assay at indicated time points following the first administration of CHS-114 for patients enrolled in SRF114-101 monotherapy dose escalation (C1D1, before drug administration/predose). A sustained increase in proinflammatory cytokines (IFN_y, TNF, CXCL10, IL-12/IL-23p40, and MIP1 β) compared to pretreatment levels across the dosing levels of CHS-114 was observed and is consistent with inducing an immune response. Data representative of 18 patients (at baseline). Error bars represent the interquartile range (IQR). Dashed red horizontal line: +25% change from baseline. All assays were significant at C2D1 relative to C1D1 with Wilcoxon test p-values < 0.05. Median values are depicted as a solid red line.

Conclusions

- CHS-114 is a novel afucosylated human IgG1 mAb that selectively and potently targets human CCR8 with no off-target binding and has the potential to overcome Treg immune suppression in the TME by depleting Treg cells and potentially increasing the T cell: Treg ratio in the tumor, enabling antitumor immunity.
- In dissociated tumor cells, CHS-114 activates NK cells and specifically induces cytotoxicity against tumor-infiltrating Tregs.
- The mechanism of action is demonstrated in a humanized CCR8 mouse tumor model: CHS-114 treatment results in significant antitumor activity and depletes human CCR8⁺ Tregs.
- CHS-114 has demonstrated an acceptable safety profile in heavily pretreated patients with advanced solid tumors, with no DLTs reported to date and TEAEs generally low grade.
- CHS-114 human PK parameters increase with dose, are approximately dose proportional, and the elimination appears linear with a half-life of about 10 days (range 9-17 days).
- Depletion of peripheral CCR8⁺ Tregs was observed in cancer patients and depletion was maintained over the dosing interval, establishing proof of mechanism.
- CHS-114 induces proinflammatory cytokines over the dosing interval, implying immune activation that is associated with depletion of CCR8⁺ Tregs.
- Preliminary results and acceptable safety profile support further evaluation of CHS-114 in combination treatment with the anti-PD-1 antibody toripalimab and other IO agents.



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